

Centrum Zastosowań Matematyki

Warsztaty

Modelowanie matematyczne i współpraca interdyscyplinarna

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Sieci Bayesowskie jako narzędzie bioinformatyka: Część I -wprowadzenie Bartek Wilczyński

Institute of Informatics, University of Warsaw

Centrum Zastosowań Matematyki, Gdańsk, 2013

Plan of the lecture

- Bayesian Network (BN) models: examples, properties, limitations, applications...
- BN models and the data: conditional probability distributions, distribution factorization, fitting to data, making predictions
- BN topology: limitations, scoring functions
- Finding optimal BNs: problem statement, complexity issues and their causes,
- Effective solutions in special cases: Dynamic BNs and BN classifiers

Models in molecular biology



Fig. 1. Modifications of the histone components of nucleosomes help regulate DNA accessibility by promoting folding or unfolding of chromatin fibers, and by recruiting chromatin remodeling complexes and other factors to specific genomic loci.

- We are facing complex systems, with many parts unknown or unobservable
- Measurements, especially of the highthroughput kind, are noisy and indirect with respect to the actual processes
- We make thousands of observations and it is vital for us to have a more concise representation of the data that we are able to interpret
- A model needs not only to be simpler than the data, but it also needs to be more general than a single dataset
- The choice of your model will always depend on the process you are interested in

Finding relationships in multi-variate data



One of the most common scenarios in "omics" projects is the one in which we look for relationships between variables in a multivariate dataset. Many statistical (regression based or qualitative) methods compete in this broad field

Advantages of probabilistic models

- Natural handling of uncertainty, both at the data and model levels
- Straightforward treatment of missing values
- Possibility to incorporate unobservable (hidden) variables
- Natural (maximum likelihood) scoring of different models for the same dataset
- Prior distributions can be used to put human knowledge into the model

Example of a simple BN



How did you feel this morning?

(great, OK, not-so-well, terrible)

Informal BN definition

- A directed acyclic graph
- with nodes representing random variables $X = \{x_i\}$
- and edges between nodes representing dependencies (very roughly, we will discuss it later)
- Each edge is directed from a parent to a child, set of parents of x is denoted by Par(x)
- Each variable is associated with a value domain and a probability distribution P(x|Par(x))

How do observations look like?

| Sex | Age | Smoking | Stress | Lung | Heart | Feel |
|-----|-------|-----------|--------|--------|--------|-------------|
| Μ | 0-20 | never | Ν | No | no | great |
| F | 70 | sometimes | Ν | minor | no | ОК |
| Μ | 50-70 | daily | Y | no | severe | Not-so-well |
| Μ | 20-50 | daily | Ν | no | minor | ОК |
| F | 70 | never | Ν | no | minor | great |
| F | 20-50 | sometimes | Y | severe | minor | Not-so-well |
| F | 20-50 | never | Y | no | no | great |
| Μ | 20-50 | sometimes | Ν | minor | no | great |
| Μ | 50-70 | never | Y | severe | no | OK |
| F | 0-20 | never | Ν | no | severe | ОК |
| Μ | 20-50 | daily | Y | no | no | ОК |
| Μ | 0-20 | daily | Ν | no | no | Not-so-well |
| Μ | 20-50 | never | Ν | minor | no | ОК |
| | | | | | | |

Conditional probability distributions

- For each variable we define the probabilities distribution in the respective domain conditional on the values of its parents
- It is assumed, that for each variable:

P(x|X) = P(x|Par(x))

| Smoking | Age | Lung=No | Lung=minor | Lung=severe |
|-----------|-------|---------|------------|-------------|
| never | 0-20 | 0.999 | 0.0009 | 0.0001 |
| sometimes | 0-20 | 0.995 | 0.004 | 0.001 |
| daily | 0-20 | 0.99 | 0.005 | 0.005 |
| never | 20-50 | 0.99 | 0.005 | 0.005 |
| sometimes | 20-50 | 0.97 | 0.02 | 0.01 |
| daily | 20-50 | 0.9 | 0.07 | 0.03 |
| | | | | |

Fitting CPDs to data

- Naturally, given enough observations, we can calculate the CPDs from contingency tables
- These contingency tables can be used to test the factorization

| Sex\Age | 0-20 | 20-50 | 50-70 | 70+ |
|---------|------|-------|-------|-----|
| Male | 55 | 43 | 42 | 25 |
| Female | 51 | 41 | 50 | 35 |

P(Age|Sex)

| Sex\Age | 0-20 | 20-50 | 50-70 | 70+ |
|---------|------|-------|-------|------|
| Male | 0.33 | 0.26 | 0.25 | 0.15 |
| Female | 0.29 | 0.23 | 0.28 | 0.20 |

Now, can we find the best graph?

- Given a dataset with observations, we can try to find the "best" network topology (i.e. the best collection of parents' sets)
- In order to solve the problem we first need to formalize our objective function to score different graphs
- A score function usually can be written as a sum over variables:

$$Score = \sum_{i=1}^{N} S_{data}(x_i, Par(x_i), Data) + S_{graph}(x_i, Par(x_i))$$

Different approaches to BN score

- There are generally 3 main approaches to defining BN scores:
 - Bayesian statistics, e.g. BDe (Herskovits *et al.*'95)
 - Information Theoretic, e.g. MDL (Lam *et al.* '94)Hypothesis testing, e.g. MMPC (Salehi *et al.* '10)
- There are also hybrid approaches, like the recent MIT (de Campos '06) approach that uses information theory and hypothesis testing

Minimum Description Length

• Graph score corresponds to the cost of encoding the CPDs:

$$g(\mathbf{Pa}) = |\mathbf{Pa}| \log n + \frac{\log N}{2} (k_X - 1) \prod_{Y \in \mathbf{Pa}} k_Y$$

• Data score corresponds to the cost of optimal encoding of the data, given the CPDs:

$$d(\mathbf{Pa}) = N \cdot H(X|\mathbf{Pa})$$

Bayesian Dirichlet equivalence

• We start from Bayes theorem:

 $P(\mathcal{G}|\mathcal{D}) \propto P(\mathcal{G})P(\mathcal{D}|\mathcal{G}) = P(\mathcal{G}) \int P(\mathcal{D}|\mathcal{G},\theta)P(\theta|\mathcal{G})d\theta$

• To write the graph score:

• And the data s

$$g(|\mathbf{Pa}|) = |\mathbf{Pa}| \log \alpha^{-1}$$

$$d(\mathbf{Pa}) = \log \left(\prod_{\mathbf{v}\in\mathcal{V}|\mathbf{Pa}|} \frac{\Gamma(\sum_{v\in\mathcal{V}}(H_{v,\mathbf{v}}+N_{v,\mathbf{v}}))}{\Gamma(\sum_{v\in\mathcal{V}}H_{v,\mathbf{v}})} \prod_{v\in\mathcal{V}} \frac{\Gamma(H_{v,\mathbf{v}})}{\Gamma(H_{v,\mathbf{v}}+N_{v,\mathbf{v}})} \right)$$

How to find the "best" network?

• We have two problems:

There are exponentially many potential parent sets
 The desired network needs to be a DAG

- It was shown by Chickering ('96) that finding an optimal BN is NP-complete
- He has shown that the problem is NP-complete even if we limit size of Parent sets to 2
- So the acyclicity criterion (2) is enough to make it practically unfeasible to find best BNs

Is it always a problem?

- If we had a situation, where the acyclicity of the network is guaranteed by an external constraint, we would only need to worry about finding the optimal parent set
- There are several practical cases when it is true. To name two:
 - Dynamic BNs
 - Classification using BNs

Dynamic Bayesian Networks

- Dynamic Bayesian
 Networks are an
 extension of the BN
 models to include
 temporal dependencies
- It is frequently used in a simple form, where only the dependencies between time points are allowed



BNs for classification

- The problem of classification, we have a number of variables, with a specified subset of *class variables*
- We are interested in models able to predict them from the other measured variables

| Sex | Age | Smoking | Stress | Lung | Heart | Feel |
|-----|-------|-----------|--------|--------|--------|-------------|
| м | 0-20 | never | N | No | no | great |
| F | 70 | sometimes | N | minor | no | OK |
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| M | 0-20 | daily | N | no | no | Not-so-well |
| м | 20-50 | never | N | minor | no | OK |
| | | | | | | |



Can we then find the solution?

Assuming that we have external constraints guaranteeing acyclicity of the network we can use the following greedy algorithm, for each variable independently:

1.
$$\mathbf{Pa} := \emptyset$$

2. for each $\mathbf{P} \subseteq \mathbf{X}'$ chosen according to $g(\mathbf{P})$

(a) if $s(\mathbf{P}) < s(\mathbf{Pa})$ then $\mathbf{Pa} := \mathbf{P}$ (b) if $g(\mathbf{P}) \ge s(\mathbf{Pa})$ then return \mathbf{Pa} ; stop

Dojer '06, Wilczynski&Dojer '09

But there are so many potential sets!

We are looking for an optimal sum of graph and data scores:

M

$$Score = \sum_{i=1}^{N} S_{data}(x_i, Par(x_i), Data) + S_{graph}(x_i, Par(x_i))$$

The graph score penalizes large parent sets. There is a limited number of parent sets with a score smaller than the optimal score. This leads to the bound on the complexity of the algorithm, in case of MDL it is:

$$\mathcal{O}(n^{\log_k N} N \log_k N)$$

Dojer '06, Wilczynski&Dojer '09

Software tools

- Banjo an MCMC method from A. Hartemink group www.cs.duke.edu/~amink/software/banjo/
- BNfinder Wilczynski&Dojer '09
- Bnlearn an R package implementing the MMPC algorithm (Salehi et al '10)
- GlobalMIT an implementation of Bnfinder greedy algorithm for the MIT score (Vinh *et al. '11)*

Summary

- BNs are a probabilistic modeling tool for multivariate data
- In general, it is NP-complete to find the optimal network (General solvers use heuristic approaches like MCMC)
- The problem lies in satisfying acyclicity of BN
- If you can avoid this problem, you can find optimal networks quickly

A few words of caution

- One needs substantial number of observations to calculate meaningful network
- The most obvious choice of the variable set is not always the best
- The fact that a network is "optimal" does not make it the "true" network
- Different scoring functions have their peculiarities, but should give you very similar "optimal" networks

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Sieci Bayesowskie jako narzędzie bioinformatyka: Część II - Zastosowania Bartek Wilczyński

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Regulatory Network Reconstruction



Dynamic Bayesian Networks

- Dynamic Bayesian
 Networks are an
 extension of the BN
 models to include
 temporal dependencies
- It is frequently used in a simple form, where only the dependencies between time points are allowed



DBNs reconstructed from "observational" expression data

- 12 microarrays (simulated)
- Optimal discretization
- No knockouts



Networks reconstructed from data with knockouts: results vary depending on number of time-points



DBN reconstruction: summary

- The results you will get will vary depending on experimental conditions
- While significant, the number of faithfully reconstructed edges will fall short of 100%
- The network will most likely not correspond to a complete (even connected) model

Modelling transcriptional networks

In Drosophila development



- Transcriptional networks as wiring diagrams
- Complexity on two different levels: many genes with many inputs



Bonn & Furlong, 2009

Transcription regulation



Genes integrate action of multiple enhancers



Wilczynski & Furlong, Dev. Biol, 2010

Temporal binding is unpredictable...





Wilczynski & Furlong, MSB, 2010

...yet reflects developmental function.



Wilczynski & Furlong, MSB, 2010

Mesoderm CRM atlas

1kb

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Multiple layers of data in transcriptional regulation



3 layer model of gene regulation

CRM Activity Model



- 8008 enhancers compiled from 15 ChIP experiments (almost 20k binding peaks)
- Activity data for ~140 enhancers divided into
 - 3 tissues (MESO, VM, SM)
 - 5 stages (4-6,7-8,9-10,1112,13-16)
- Gene expression data for 5082 genes from the BDGP database

Model structure and optimization





Model structure and optimization



Expectation-Maximization approach



- CRM activity is in a large part a hidden variable for us
- With a probabilistic model we can use iterative strategy to find the optimal model combining the information from both TF binding and gene expression levels
- In the E-step we estimate the CRM activity and then in the M-step we maximize the BN and promoter models

Gene expression prediction



- Using known enhancers from CAD and in-situ annotations from BDGP
- We are trying to train a model predicting expression of target genes in different tissues
- Target gene assignment is key to linking CRM activity with expression of Targets

EM algorithm (1)



STEP 0: initialize parameter values to some (usually informed) guess In our case: learn Θ_1 from CAD and set Θ_2 to maximal value.

EM algorithm (2)



Once we have parameter values, we can calculate expected probabilities for hidden variables. That's E-STEP (expectation).

EM algorithm (3)



Once we have the estimated probabilities of hidden variables, we can find new set of parameters maximizing the likelihood function given our current expectation. That's M-STEP (maximization)

EM algorithm (4)



We can iterate these two steps until we reach convergence. In the end, we have a full model which allows us to "predict" C from A.

Validation of predictions by in-situ hybrydization



Wilczynski et al. submitted

Summary

- Integrated model gives better predictions of gene expression and enhancer activity
- We see indication of enhancer sharing among developmental genes
- Very long range interactions do happen in Drosophila (typical locus is 50-100kb =~ 10 genes)
- We can find "missing players" that should be specific to related tissues

Histone modifications and their role in transcription



The figure illustrates nucleosome models and major posttranslational modifications which play essential roles in gene expression regulation and disease processes







Nature Reviews | Cancer

Modencode - Histone code?

| Chromatin States | | | | Histone Marks - enrichment | | | | | | | | | | | ne | | | | | | |
|-------------------------------|-----------|-------------------------|----------|----------------------------|----------|----------|---------|---------|--------|----------|-----------|----------|---------|---------|--------------|--------------|----------------------|---------|---------|----------|----------|
| | | | | | | | | | | <u> </u> | none high | | | | | 2 | | | | | |
| State Annotation Summary | Discrete | Continuous Intensity | H3K36me3 | H3K79me1 | H2B-ubiq | H3K79me2 | H3K4me2 | H3K4me3 | H3K9ac | H4K16ac | H3K4me1 | H3K36me1 | H3K18ac | H3K27ac | H1 depletion | H4 depletion | H3K23ac depletion | H3K9me3 | H3K9me2 | H3K27me3 | % of gen |
| | d1 | | 1 | 0 | 1 | 3 | 47 | 92 | 57 | 7 | 0 | 0 | 0 | 3 | 1 | 12 | 7 | 0 | 0 | 0 | 2.09 |
| | d2 | | 95 | 20 | 10 | 10 | 79 | 93 | 24 | 9 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0.26 |
| Active TSS/exon | d3 | c1 | 52 | 3 | 55 | 79 | 99 | 100 | 92 | 45 | 7 | 0 | 1 | 13 | 1 | 2 | 1 | 0 | 0 | 0 | 1.77 |
| | <u>d4</u> | Ŭ., | 57 | 22 | 73 | 77 | 93 | 64 | 5 | 7 | 23 | 1 | 0 | 4 | 0 | 1 | 0 | 1 | 1 | 0 | 1.45 |
| | d5 | | 2 | 0 | 8 | 11 | 78 | 87 | 92 | 39 | 4 | 1 | 59 | 89 | 4 | 22 | 3 | 0 | 0 | 0 | 1.10 |
| | <u>ae</u> | | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 1 | 1 | 24 | 11 | 0 | 0 | 0 | 2.95 |
| | 40 | | /5 | 44 | 88 | 5/ | 0 | 1 | 0 | - 1 | 4 | 0 | 1 | 1 | 0 | 2 | 0 | 0 | 0 | 0 | 2.39 |
| Active exon, elongation | <u>d0</u> | c2 | 72 | 67 | 54 | 14 | 1 | 0 | 0 | | | 1 | 0 | 0 | 0 | 5 | 26 | 50 | 77 | 0 | 0.95 |
| | d10 | | 13 | 37 | 2/1 | 14 | | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 0 | | 20 | 00 | 1 | 1 | 3.00 |
| | d11 | | 2 | 2 | 7 | 14 | 63 | 7 | 64 | 82 | 84 | 46 | 98 | 98 | 4 | 12 | 0 | 0 | 0 | 0 | 1.56 |
| | d12 | c 3 | 2 | 2 | 88 | 69 | 79 | 2 | 47 | 23 | 55 | 87 | 84 | 59 | 0 | 3 | 0 | 0 | 0 | 0 | 0.78 |
| | d13 | | 4 | 1 | 79 | 73 | 100 | 94 | 87 | 32 | 24 | 66 | 83 | 73 | 1 | 5 | 0 | 0 | 0 | 0 | 0.50 |
| Active intron, enhancer | d14 | | 3 | 1 | 1 | 1 | 13 | 0 | 15 | 11 | 42 | 2 | 56 | 75 | 1 | 11 | 1 | 0 | 0 | 0 | 1.24 |
| | d15 | | 0 | 8 | 3 | 17 | 8 | 0 | 13 | 15 | 63 | 93 | 81 | 26 | 0 | 3 | 0 | 0 | 0 | 1 | 1.44 |
| | d16 | 1 1 | 0 | 5 | 88 | 64 | 3 | 0 | 30 | 3 | 15 | 95 | 84 | 3 | 0 | 1 | 0 | 0 | 0 | 0 | 0.48 |
| | d17 | | 3 | 2 | 4 | - 4 | 92 | 12 | 13 | 2 | 33 | 12 | 9 | 11 | 1 | - 4 | 1 | 1 | 0 | 13 | 0.64 |
| | d18 | | 0 | 10 | 2 | 2 | 0 | 0 | 3 | 1 | 6 | 88 | 21 | 0 | 0 | 1 | 0 | 0 | 0 | 7 | 1.66 |
| Open chromatin | d19 | ~1 | 0 | 15 | 84 | 36 | 2 | 0 | 3 | 3 | 7 | 72 | - 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1.22 |
| open chromaun | d20 | 64 | 1 | 4 | 0 | 1 | 1 | 0 | 3 | 1 | 89 | 11 | 10 | 2 | 0 | 5 | 0 | 1 | 0 | 3 | 2.07 |
| | d21 | | 0 | 0 | 0 | 0 | 0 | 0 | 5 | 0 | 1 | 3 | 7 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2.53 |
| | d22 | | 1 | 1 | 0 | 0 | 1 | 0 | 0 | 78 | 3 | 2 | 1 | 0 | 0 | 2 | 0 | 1 | 3 | 1 | 2.21 |
| Male X genes (DC), exon | d23 | c5 | 65 | 21 | 29 | 5 | 3 | 1 | 0 | 99 | 5 | 1 | 1 | 1 | 0 | - 4 | 1 | 1 | 2 | 0 | 0.98 |
| | d24 | | 28 | 5 | 21 | 11 | 88 | 58 | 26 | 98 | 11 | 1 | 2 | 7 | 0 | 3 | 0 | 0 | 0 | 0 | 1.22 |
| Polycomb | d25 | C6 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 81 | 5.58 |
| Heterochromatin | d26 | c7 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 4 | 96 | 82 | 91 | 0 | 2.26 |
| | d27 | | 61 | 5 | 4 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 19 | 82 | 55 | 35 | 0 | 0.84 |
| Heterochromatin-like in euch | 028 | c8 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 3 | 91 | 25 | 2 | 1.62 |
| Pool internenie ouebremetin | 029 | <u></u> | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 4 | 1 | 35 | 0 | 2.52 |
| Dasal, intergenic euchromatin | u30 | 69 | 0 | U | U | U | U | U | U | U | U | 0 | U | U | U | U | U | 0 | U | U | 50.75 |

Modencode, Roy et al, Science 2010

Getting a clear picture - BiTS-Chip



Alexa488 (Log)

Bonn et al. Nat. Genet, 2012

ppa

What do we see in regions with known genes



md Mef2

Can we extract patterns for "known" enhancers?



Can we model the relationship between histone marks and activity?



How does the network work?



Expressed In Mesoderm at 6-8h



How can it be useful?

Scoring the Intergenic Genome



Bonn et al. Nat. Genet, 201

Predicted Enhancers

Does it work?



- 12 positive and 4 negative predictions tested
- >90% success!

New projects: Insulators and their interactions



- Chromosomes are packed in the nucleus in a nonrandom fashion
- Chromosomal interactions are mediated by proteins, including insulators

Playing with chromosome interaction data



- Hi-C and ChIA-Pet protocols allow for measurement of chromatin interactions
- Getting this type of data is still difficult
- We can use computational methods to predict interactions

Data from Handoko et al. 2011

Using histone modification similarity to find likely pairs



Histone modification profiles match between interacting anchor points

Loop center

If we define a reasonable measure, we can see this signal on a genome-wide scale to be significant



Making predictions

- We can use MCMC algorithm to search the (HUGE!) space of possible interaction ensembles (not real 3D conformations)
- We can get accuracy of ~50% on a small set of interactions
- That is significant, but not quite satisfactory
- Predicting interaction dynamics is feasible





Predicting boundary elements from modEncode data



- We can use all chromatin IP data available in modENCODE for late embryos and try to predict domain boundaries
- We will use the Hi-C derived data as our main interaction set and Dam-ID derived data as additional validation

BN classifier can predict boundaries

- Using BN classifiers trained on modENCODE data, we can predict position of boundary elements
- This method outperforms HMMs and clustering of histone modification data



Predictions make sense, and the model brings new information



DamID validation results

Impact of signals

Some predictions are unexpected



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- Charles Girardot
- Robert Zinzen
- Stefan Bonn





Post-doc positions available



